

1. Li, W. *et al.* Evaluation of the safety and efficiency of cytotoxic T cell therapy sensitized by tumor antigens original from T-ALL-iPSC in vivo. *Cancer Innovation* **3**, e95 (2024) doi: [10.1002/cai2.95](https://doi.org/10.1002/cai2.95).
2. Yasui, H. *et al.* Immune Function of Chimeric Antigen Receptor T Cells Quantitatively Assessed Via Molecular Imaging Flow Cytometry. *Blood* **142**, 6821 (2023) doi: [10.1182/blood-2023-173065](https://doi.org/10.1182/blood-2023-173065).
3. Jiang, W. *et al.* Novel mesothelin-targeted chimeric antigen receptor-modified UNKT cells are highly effective in inhibiting tumor progression. *Pharmacological Research* **197**, 106942 (2023) doi: [10.1016/j.phrs.2023.106942](https://doi.org/10.1016/j.phrs.2023.106942).
4. Hasebe, Y. & Naoe, M. Gamma-delta T cells are optimal immune cell carrier vehicles for adenovirus vector-based gene therapy. *The Showa University Journal of Medical Sciences* **35**, 103–111 (2023) doi: [10.15369/sujms.35.103](https://doi.org/10.15369/sujms.35.103).
5. Zhang, G., Liao, Y., Pan, X. & Zhang, X. Exosomes from HPV-16 E7-pulsed dendritic cells prevent the migration, M1 polarization, and inflammation of macrophages in cervical cancer by regulating catalase 2 (CAT2). *Annals of Translational Medicine; Vol 10, No 4 (February 28, 2022): Annals of Translational Medicine* (2022) doi: [10.21037/atm-21-6998](https://doi.org/10.21037/atm-21-6998).
6. Tsuchiya, Y., Kobayashi, H., Kanno, H. & Yamamoto, T. Beta-Tricalcium Phosphate as a Possible Adjuvant in $\gamma\delta$ T Cell-Based Immune Therapy for Human Disorders. *Tokyo Women's Medical University Journal* **6**, 101–107 (2022) doi: [10.24488/twmuj.2022012](https://doi.org/10.24488/twmuj.2022012).
7. Ono, S. *et al.* Trametinib improves Treg selectivity of anti-CCR4 antibody by regulating CCR4 expression in CTLs in oral squamous cell carcinoma. *Scientific Reports* **12**, 21678 (2022) doi: [10.1038/s41598-022-22773-1](https://doi.org/10.1038/s41598-022-22773-1).
8. Li, Z. *et al.* Development of the T-ALLiPSC-based therapeutic cancer vaccines for T-cell acute lymphoblastic leukemia. *Medical Oncology* **39**, 200 (2022) doi: [10.1007/s12032-022-01809-6](https://doi.org/10.1007/s12032-022-01809-6).
9. Tomogane, M. *et al.* Human V γ 9V δ 2 T cells exert anti-tumor activity independently of PD-L1 expression in tumor cells. *Biochemical and Biophysical Research Communications* **573**, 132–139 (2021) doi: [10.1016/j.bbrc.2021.08.005](https://doi.org/10.1016/j.bbrc.2021.08.005).
10. Takahashi, N. *et al.* Construction of in vitro patient-derived tumor models to evaluate anticancer agents and cancer immunotherapy. *Oncol Lett* **21**, 406 (2021) doi: [10.3892/ol.2021.12667](https://doi.org/10.3892/ol.2021.12667).
11. Kondo, Y. *et al.* Improving function of cytotoxic T-lymphocytes by transforming growth factor- β inhibitor in oral squamous cell carcinoma. *Cancer Science* **112**, 4037–4049 (2021) doi: [10.1111/cas.15081](https://doi.org/10.1111/cas.15081).

12. Zeng, G. *et al.* Cryopreservation of peripheral blood mononuclear cells using uncontrolled rate freezing. *Cell and Tissue Banking* **21**, 631–641 (2020) doi: [10.1007/s10561-020-09857-w](https://doi.org/10.1007/s10561-020-09857-w).
13. Sasawatari, S., Okamoto, Y., Kumanogoh, A. & Toyofuku, T. Blockade of N-Glycosylation Promotes Antitumor Immune Response of T Cells. *J. Immunol.* **204**, 1373 (2020) doi: [10.4049/jimmunol.1900937](https://doi.org/10.4049/jimmunol.1900937).
14. Noguchi, J. *et al.* Successful activation of rat T lymphocytes by sperm specific antigens in vitro. *Journal of Reproduction and Development* **66**, 599–605 (2020) doi: [10.1262/jrd.2020-106](https://doi.org/10.1262/jrd.2020-106).
15. Murakami, S. *et al.* Combining T-cell-based immunotherapy with venetoclax elicits synergistic cytotoxicity to B-cell lines in vitro. *Hematological Oncology* **38**, 705–714 (2020) doi: [10.1002/hon.2794](https://doi.org/10.1002/hon.2794).
16. Miyashita, M. *et al.* Sphere-derived Prostate Cancer Stem Cells Are Resistant to $\gamma\delta$ T Cell Cytotoxicity. *Anticancer Res* **40**, 5481–5487 (2020) doi: [10.21873/anticancer.14559](https://doi.org/10.21873/anticancer.14559).
17. Koyama, I. *et al.* A Clinical Trial With Adoptive Transfer of Ex Vivo-induced, Donor-specific Immune-regulatory Cells in Kidney Transplantation—A Second Report. *Transplantation* **104**, (2020) doi: [10.1097/TP.0000000000003149](https://doi.org/10.1097/TP.0000000000003149).
18. Futami, M. *et al.* The novel multi-cytokine inhibitor TO-207 specifically inhibits pro-inflammatory cytokine secretion in monocytes without affecting the killing ability of CAR T cells. *PLOS ONE* **15**, e0231896 (2020) doi: [10.1371/journal.pone.0231896](https://doi.org/10.1371/journal.pone.0231896).
19. Fujiwara, K. *et al.* Predicting the Efficacy and Safety of TACTICs (Tumor Angiogenesis-Specific CAR-T Cells Impacting Cancers) Therapy for Soft Tissue Sarcoma Patients. *Cancers* **12**, (2020) doi: [10.3390/cancers12102735](https://doi.org/10.3390/cancers12102735).
20. Abe, Y. *et al.* Application of gamma-delta T cells obtained by ascites filtration for immunotherapy against malignant refractory ascites. *Annals of Cancer Research and Therapy* **27**, 73–79 (2019) doi: [10.4993/acrt.27.73](https://doi.org/10.4993/acrt.27.73).
21. Shimizu, T., Tomogane, M., Miyashita, M., Ukimura, O. & Ashihara, E. Low dose gemcitabine increases the cytotoxicity of human V γ 9V δ 2 T cells in bladder cancer cells in vitro and in an orthotopic xenograft model. *null* **7**, e1424671 (2018) doi: [10.1080/2162402X.2018.1424671](https://doi.org/10.1080/2162402X.2018.1424671).
22. Kimura, H., Matsui, Y., Ishikawa, A., Nakajima, T. & Iizasa, T. Randomized controlled phase III trial of adjuvant chemoimmunotherapy with activated cytotoxic T cells and dendritic cells from regional lymph nodes of patients with lung cancer. *Cancer Immunology, Immunotherapy* **67**, 1231–1238 (2018) doi: [10.1007/s00262-018-2180-6](https://doi.org/10.1007/s00262-018-2180-6).

23. Abe, Y., Kobayashi, H., Akizawa, Y., Ishitani, K. & Matsui, H. Role of IL-18 on the activation of V δ 2⁺ T cells – For the development of novel cancer immunotherapy –. *Annals of Cancer Research and Therapy* **26**, 71–76 (2018) doi: [10.4993/acrt.26.71](https://doi.org/10.4993/acrt.26.71).
24. Abe, Y. *et al.* Possible Application of Ascites-infiltrating Gamma-delta T Cells for Adoptive Immunotherapy. *Anticancer Res* **38**, 4327–4331 (2018) doi: [10.21873/anticancer.12732](https://doi.org/10.21873/anticancer.12732).
25. Takagi, M. *et al.* Haploinsufficiency of TNFAIP3 (A20) by germline mutation is involved in autoimmune lymphoproliferative syndrome. *Journal of Allergy and Clinical Immunology* **139**, 1914–1922 (2017) doi: [10.1016/j.jaci.2016.09.038](https://doi.org/10.1016/j.jaci.2016.09.038).
26. Nishio-Nagai, M., Suzuki, S., Yoshikawa, K., Ueda, R. & Kazaoka, Y. Adoptive immunotherapy combined with FP treatment for head and neck cancer: An in vitro study. *Int J Oncol* **51**, 1471–1481 (2017) doi: [10.3892/ijo.2017.4142](https://doi.org/10.3892/ijo.2017.4142).
27. Todo, S. *et al.* A pilot study of operational tolerance with a regulatory T-cell-based cell therapy in living donor liver transplantation. *Hepatology* **64**, 632–643 (2016) doi: [10.1002/hep.28459](https://doi.org/10.1002/hep.28459).
28. Inoo, K. *et al.* Immunological quality and performance of tumor vessel-targeting CAR-T cells prepared by mRNA-EP for clinical research. *Molecular Therapy - Oncolytics* **3**, 16024 (2016) doi: [10.1038/mto.2016.24](https://doi.org/10.1038/mto.2016.24).
29. Eom, H.-S. *et al.* Phase I Clinical Trial of 4-1BB-based Adoptive T-Cell Therapy for Epstein-Barr Virus (EBV)-positive Tumors. *J Immunother* **39**, 140–148 (2016) doi: [10.1097/CJI.000000000000113](https://doi.org/10.1097/CJI.000000000000113).
30. Liu, H. *et al.* Comparative study of different procedures for the separation of peripheral blood mononuclear cells in cytokine-induced killer cell immunotherapy for hepatocarcinoma. *Tumor Biology* **36**, 2299–2307 (2015) doi: [10.1007/s13277-014-2837-5](https://doi.org/10.1007/s13277-014-2837-5).
31. Kimura, H. *et al.* Randomized controlled phase III trial of adjuvant chemo-immunotherapy with activated killer T cells and dendritic cells in patients with resected primary lung cancer. *Cancer Immunology, Immunotherapy* **64**, 51–59 (2015) doi: [10.1007/s00262-014-1613-0](https://doi.org/10.1007/s00262-014-1613-0).
32. Narita, T. *et al.* HTLV-1 bZIP Factor–Specific CD4 T Cell Responses in Adult T Cell Leukemia/Lymphoma Patients after Allogeneic Hematopoietic Stem Cell Transplantation. *J. Immunol.* **192**, 940 (2014) doi: [10.4049/jimmunol.1301952](https://doi.org/10.4049/jimmunol.1301952).
33. Morisaki, T. *et al.* NKG2D-directed Cytokine-activated Killer Lymphocyte Therapy Combined with Gemcitabine for Patients with Chemoresistant Metastatic Solid Tumors. *Anticancer Res* **34**, 4529 (2014).
34. Masaki, A. *et al.* Autologous Tax-Specific CTL Therapy in a Primary Adult T Cell Leukemia/Lymphoma Cell–Bearing NOD/Shi-*scid*, IL-2R^{null} Mouse Model. *J. Immunol.* **191**, 135 (2013) doi: [10.4049/jimmunol.1202692](https://doi.org/10.4049/jimmunol.1202692).

35. Suzuki, S. *et al.* Tax is a potential molecular target for immunotherapy of adult T-cell leukemia/lymphoma. *Cancer Science* **103**, 1764–1773 (2012) doi: [10.1111/j.1349-7006.2012.02371.x](https://doi.org/10.1111/j.1349-7006.2012.02371.x).
36. Tsuda, J. *et al.* Involvement of CD56^{bright}CD11c⁺ Cells in IL-18–Mediated Expansion of Human $\gamma\delta$ T Cells. *J. Immunol.* **186**, 2003 (2011) doi: [10.4049/jimmunol.1001919](https://doi.org/10.4049/jimmunol.1001919).
37. Kobayashi, H., Tanaka, Y., Yagi, J., Minato, N. & Tanabe, K. Phase I/II study of adoptive transfer of $\gamma\delta$ T cells in combination with zoledronic acid and IL-2 to patients with advanced renal cell carcinoma. *Cancer Immunology, Immunotherapy* **60**, 1075–1084 (2011) doi: [10.1007/s00262-011-1021-7](https://doi.org/10.1007/s00262-011-1021-7).
38. Suzuki, T. *et al.* The antitumour effect of $\{\gamma\delta\}$ T-cells is enhanced by valproic acid-induced up-regulation of NKG2D ligands. *Anticancer Res* **30**, 4509–4513 (2010).
39. Li, W. *et al.* Effect of IL-18 on Expansion of $\gamma\delta$ T Cells Stimulated by Zoledronate and IL-2. *Journal of Immunotherapy* **33**, 287–296 (2010) doi: [10.1097/CJI.0b013e3181c80ffa](https://doi.org/10.1097/CJI.0b013e3181c80ffa).
40. Watanabe, K. *et al.* CD137-guided isolation and expansion of antigen-specific CD8 cells for potential use in adoptive immunotherapy. *International Journal of Hematology* **88**, 311–320 (2008) doi: [10.1007/s12185-008-0134-z](https://doi.org/10.1007/s12185-008-0134-z).

ALyS 705

1. Suematsu, M. *et al.* PiggyBac Transposon-Mediated CD19 Chimeric Antigen Receptor-T Cells Derived From CD45RA-Positive Peripheral Blood Mononuclear Cells Possess Potent and Sustained Antileukemic Function. *Frontiers in Immunology* **13**, (2022) doi: doi.org/10.3389/fimmu.2022.770132.
2. Yagyu, S. *et al.* A lymphodepleted non-human primate model for the assessment of acute on-target and off-tumor toxicity of human chimeric antigen receptor-T cells. *Clinical & Translational Immunology* **10**, e1291 (2021) doi: [10.1002/cti2.1291](https://doi.org/10.1002/cti2.1291).
3. Tomida, A. *et al.* Inhibition of MEK pathway enhances the antitumor efficacy of chimeric antigen receptor T cells against neuroblastoma. *Cancer Sci* **112**, 4026–4036 (2021) doi: [10.1111/cas.15074](https://doi.org/10.1111/cas.15074).
4. Nakamura, K. *et al.* Autologous antigen-presenting cells efficiently expand piggyBac transposon CAR-T cells with predominant memory phenotype. *Molecular Therapy - Methods & Clinical Development* **21**, 315–324 (2021) doi: [10.1016/j.omtm.2021.03.011](https://doi.org/10.1016/j.omtm.2021.03.011).
5. Kubo, H. *et al.* Development of non-viral, ligand-dependent, EPHB4-specific chimeric antigen receptor T cells for treatment of rhabdomyosarcoma. *Molecular Therapy - Oncolytics* **20**, 646–658 (2021) doi: [10.1016/j.omto.2021.03.001](https://doi.org/10.1016/j.omto.2021.03.001).

6. Morokawa, H. *et al.* Autologous non-human primate model for safety assessment of piggyBac transposon-mediated chimeric antigen receptor T cells on granulocyte–macrophage colony-stimulating factor receptor. *Clinical & Translational Immunology* **9**, e1207 (2020) doi: [10.1002/cti2.1207](https://doi.org/10.1002/cti2.1207).
7. Mie, K. *et al.* Influence of transfusion of lymphokine-activated T killer cells on inflammatory responses in dogs after laparotomy. *J Vet Med Sci* **78**, 579–585 (2016) doi: [10.1292/jvms.15-0626](https://doi.org/10.1292/jvms.15-0626).
8. Mie, K., Shimada, T., Akiyoshi, H., Hayashi, A. & Ohashi, F. Change in peripheral blood lymphocyte count in dogs following adoptive immunotherapy using lymphokine-activated T killer cells combined with palliative tumor resection. *Veterinary Immunology and Immunopathology* **177**, 58–63 (2016) doi: [10.1016/j.vetimm.2016.06.007](https://doi.org/10.1016/j.vetimm.2016.06.007).